

structural features of the tyrosines (phenoxy ring rotation angle) or with the values of the parameters obtained from the DFT calculation. This allows identification of the tyrosine in the protein responsible for the observed radical.

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Archaeal Protoglobin Structures: Novel Ligand Diffusion Paths And Heme Reactivity Modulation

Marco Nardini¹, Alessandra Pesce², Paolo Ascenzi³, Massimo Coletta⁴,

Martino Bolognesi^{1,5}.

¹Univ. of Milano, Milano, Italy, ²Univ. of Genova, Genova, Italy, ³Univ. of Roma Tre, Rome, Italy, ⁴Univ. of Roma Tor Vergata, Rome, Italy,

⁵Dept. Biomol. Sciences and Biotechnology - University of Milano, Milano, Italy.

Despite its strict anaerobic nature, *M. acetivorans* genome hosts genes related to O₂ metabolism; among these, an open reading frame encodes for a "protoglobin" (NP_617780; Pgb). Pgb is a single domain heme protein of ~195 amino acids, related to the N-terminal domain of archaeal and bacterial globin coupled sensor proteins (GCS; Freitas et al. (2004) Proc. Natl. Acad. Sci. USA 101, 6675-6680). Sequence comparisons indicate that Pgb, despite their 30-35% larger size, are structurally related to single chain hemoglobins (about 150 amino acids, 3-on-3 α -helical sandwich, 12-16% residue identity to Pgb), and to the heme-based aerotaxis transducer sensor domain of *Bacillus subtilis* GCS. Pgb binds O₂, CO, and NO reversibly in vitro. Functional and evolutionary issues are openly debated: Pgb may facilitate O₂ detoxification in vivo promoting electron transfer to O₂, or may act as CO sensor/supplier in methanogenesis.

Our previous studies unravelled the 3D structure of *M. acetivorans* Pgb, and of its ligand binding properties (Nardini et al. (2008) EMBO Reports 9, 157-163). We showed that Ma-Pgb heme- domain is strongly related in tertiary and quaternary structure to the N-terminal domain of archaeal and bacterial GCSs. Contrary to known globins, however, Pgb-specific loops and a N-terminal extension completely bury the heme within the protein matrix. A new access route to the heme, built by two Pgb-specific apolar tunnels reaching the heme distal site from locations at the B/G and B/E helix interfaces was highlighted. We present here structural and ligand binding properties of four Ma-Pgb mutants (at sites B10, B12, G8, G11) that were designed in order to probe the role of the heme access tunnels previously described. The atomic resolution structures will be discussed at the light of the kinetic parameters measured for the mutant Ma-Pgb.

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Structural Probes Of Reactive Intermediates Of Dehaloperoxidase From *Amphitrite ornata*

Stefan Franzen, Vesna de Serrano, Michael F. Davis, Matt Thompson.

NC State University, Raleigh, NC, USA.

The enzyme dehaloperoxidase (DHP) from the marine worm *Amphitrite ornata* is a unique hemoglobin that functions as a peroxidase, capable of converting 2,4,6-trihalo-phenols (TBP, TCP, and TFP) into the corresponding 2,6-dihalogenated quinones as well as other products. In this overview talk we discuss the structure and function of DHP using X-ray crystallography and nuclear magnetic resonance (NMR) to discuss the large differences between DHP function and hemoglobin function despite the strong structural similarities. The position of halogenated phenols inside the distal pocket is one anomalous feature of DHP that is not observed in any other hemoglobin or myoglobin. The X-ray crystal structure of DHP reveals that the distal histidine is flexible and has two major conformations. The closed conformation (named by analogy with sperm whale myoglobin) is enforced by the binding of a sixth ligand to the heme iron. In the open conformation, observed in the deoxy DHP X-ray crystal structure, the distal histidine is in a solvent exposed conformation. The role of the histidine in coupling the binding of substrate and inhibitors will be discussed in the context of the mechanism for formation of compound I, compound II and a novel intermediate called compound RH that appears to be crucial to the cycling of DHP between hemoglobin and peroxidase function. The NMR data reveal that there are both interior and exterior binding sites for the substrate. This aspect will be discussed along with evidence from optical and EPR spectroscopy to understand the electron transfer kinetics of DHP.

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Structural Analysis of Hemoglobins and Myoglobins Using MD Simulations

Mariano A. Scorciapino¹, Arturo Robertazzi¹, Enrico Spiga¹,

Mariano Casu¹, Paolo Ruggerone^{1,2}, Matteo Ceccarelli^{1,2}.

¹University of Cagliari, Monserrato, Italy, ²CNR-SLACS, Cagliari, Italy.

In the family of respiratory proteins, hemoglobins and myoglobins have been the first to be crystallized in X-ray. Despite their precise 3D structures are available at high resolution, some questions regarding the microscopic functioning remain yet open. The R to T switching mechanism in hemoglobins and the ligand escape process in myoglobins remain still under debate.

Thanks to the small size, myoglobin is the preferred candidate also for the more general structure-function paradigm. In the interior of myoglobin five main docking sites have been identified, especially with X-ray NMR, and for long time these Xenon cavities have been classified as packing defects. Recently, it was shown that they might be involved in ligands migration path, even if mechanisms used by myoglobin to connect these cavities is still unknown as well as processes regulating its biologic functions. In this work we made use of standard MD simulations of solvated myoglobin to characterize internal cavities. Our principal results is that we have found several secondary cavities showing volume and occurrence at least comparable to that of Xenon cavities. In order to rationalize and in-depth analyze such a huge amount of data (ca. 30000 cavities/10 ns MD), special cluster-analysis was applied: we classified all cavities with respect to the position, size and occurrence as function of simulation time ascribing them to different clusters. This analysis implicitly highlights possible ligand migration paths for small ligands within the protein matrix allowing to quantitatively compare dynamical behaviour of different myoglobins towards different ligands. Our suggestion that the secondary cavities constitute the preferred path for ligand escape is also supported by explicit metadynamics simulations of ligand escape.

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Auto-Oxidation of Human Hemoglobin and the Roles of Distal Heme Pocket Substitutions

Natalie Weir, David H. Mailett, Tong-Jian Shen, Chien Ho.

Carnegie Mellon University, Pittsburgh, PA, USA.

This study investigates the auto-oxidation reaction of human normal adult hemoglobin (Hb A) and the effects of distal heme pocket substitutions, P₅₀, and tetramer-dimer dissociation on the rate and mechanism of auto-oxidation. Recombinant hemoglobins (rHbs) with single amino acid substitutions at helical positions E11 and B10 have been expressed in *Escherichia coli* and purified, as well as di- α linked and octameric rHbs. These rHbs include: rHb (α V62L), rHb (α V62I), rHb (β V67L), rHb (β V67I), rHb (α L29W), rHb (α L29F), rHb (α -Gly- α / β gene di- α / β), rHb (di α L29F), rHb (di α L29W), and rHb (β G83C). Auto-oxidation measurements were conducted with 32 μ M heme in MES buffer (pH 6.5) for 60 hours at 35 °C. A monophasic nature of auto-oxidation has been observed for Hb A and a biphasic nature for all other rHbs. In comparison to the other mutants, including di- α linked and octameric rHbs, rHb (α L29F) is most resistant to oxidation and rHb (α L29W) is the least resistant to oxidation. Characterization of three novel rHbs; (β L28F, β V67I), (α L29F, β V67I), (α L29F, α V62I) will test whether the fast and slow phases of the observed biphasic nature of auto-oxidation can be attributed to the mutated and wild-type subunits, respectively. Our studies will also provide new insights into the roles of amino acid residues in the distal heme pockets on the structure-function relationship in hemoglobin (Supported by NIH grants HL-024525 and GM-084614, HHMI, and The Arnold and Mabelle Beckman Undergraduate Research Scholars Program).

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The Use Of Glassy Matrices To Identify Intermediates In The Nitric Oxide Dioxigenase Reaction Of Hemoglobins And Myoglobins

Mahantesh S. Navati, Joel M. Friedman.

Albert Einstein College Of Medicine, Bronx, NY, USA.

The nitric oxide dioxigenase (NOD) reaction of nitric oxide with ferrous oxy derivatives of Hb and Mb is important from many physiological standpoints. Several mechanisms have been proposed; however, the experimental difficulties in probing this reaction in solution at physiological pH and ambient temperatures have precluded an unambiguous determination of the sequence and nature of intermediates. We have developed a method of following the progression of this reaction in glassy matrices that allows for the trapping and probing of key intermediates. The technique is based on incorporation of O₂ derivatives of Hb and Mb in a thin glassy matrix (derived from trehalose) that lines the inner wall of an optical quality tube. After purging the sample with dry nitrogen to remove the unbound excess oxygen, the tube is filled with NO. Absorption spectroscopy is used to follow the spectral progression initiated as the NO slowly accesses the heme bound oxygen. The spectra reveal an intermediate that resembles the spectrum attributed to the bound peroxynitrite intermediate. The final product under these conditions is a species with a spectrum that is identical to that which is generated when met-Hb(Mb) is incorporated into a glass in the presence of an excess of nitrate. The spectrum attributed to the nitrate

product only appears when the glass is dry. The results are consistent with water being an effective competitor for the ferric heme site in the presence of nitrate.

Platform BA: Calcium Signaling Pathways

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Isoform-specific Regulation Of The Ca-sensitive Transcription Factor NFAT In The Cardiovascular System

Andreas Rinne¹, Kathrin Banach², Donald M. Bers³, Lothar A. Blatter¹.

¹Rush University, Chicago, IL, USA, ²UIC, Chicago, IL, USA, ³UC Davis, Davis, CA, USA.

NFAT transcription factors (Nuclear Factor of Activated T-cells) mediate Ca-sensitive gene transcription and are involved in cardiovascular remodelling. Nuclear localization of NFAT is dynamically regulated by intracellular Ca signals yielding to dephosphorylation and nuclear translocation of NFAT, and activity of intracellular kinases that induce nuclear export. The aim of this study was to analyze the regulation of NFAT in vascular endothelial cells and adult cardiomyocytes. Subcellular distribution of NFAT-GFP fusion proteins (isoforms NFATc1 and NFATc3) was analyzed with confocal microscopy and intracellular Ca ([Ca]_i) was measured simultaneously using rhod-2. In calf pulmonary arterial endothelial (CPAE) cells, application of ATP (5 μM) induced nuclear localization of both isoforms (quantified as an increase in NFAT_{NUC}/NFAT_{CYT} ratio). Subsequent attenuation of [Ca]_i to facilitate nuclear export resulted in substantial export of NFATc3 to the cytoplasm, which was sensitive to Leptomycin B (40 nM). Previously translocated NFATc1 was only partially affected by nuclear export, indicating isoform-specific regulation of NFAT in endothelial cells.

In cardiac myocytes regulation of NFAT was isoform-, and tissue-specific: NFATc1 displayed nuclear localization in quiescent myocytes, which was dependent on [Ca]_i and further enhanced by blocking nuclear export (Leptomycin B) or by inhibition of intracellular kinases (20 mM LiCl, 1 μM alsterpaullone or 1 μM SP600125). In contrast, NFATc3 was distributed in the cytoplasm of quiescent cells. Incubation with Leptomycin B, but not inhibition of nuclear kinases induced nuclear localization of NFATc3 in ventricular cells. Incubation with the G_q protein-coupled receptor agonists endothelin-1 (100 nM) and Ang II (2 μM) induced nuclear localization of NFATc3 only in atrial, but not ventricular cells. We conclude that (i) regulation of nuclear NFAT in the cardiovascular system is isoform- and tissue specific and (ii) dynamically regulated by activity of nuclear export pathways.

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Alterations In Binding Properties Of Myocardial Nuclear Membrane Receptors Induce Nuclear Calcium Overload In Rat Ischemia-reperfusion

Huamei He¹, Lezhi Zhang², Ailing Fu², Hong Zhang², Bin Li²,

Aiping Tang², Zhou Hong².

¹Children's Hospital Boston and Harvard Medical School, Boston, MA,

USA, ²Department of Pharmacology, Third Military Medical University, Chongqing, China.

Cell nuclei possess an independent calcium regulatory system consisting of nuclear Ca²⁺-ATPases (NCA), IP₃ receptors (IP₃R), IP₄ receptors (IP₄R), ryanodine receptors (RyR), and nuclear pore complexes (NPC). We studied the changes in Ca²⁺_n and its regulatory system in rat model of myocardial ischemia-reperfusion injury (IRI) induced by 30 min coronary occlusion followed by 180 min reperfusion. The Ca²⁺_n content in isolated nuclei was measured with atomic absorption spectrophotometer. NPC permeability was assessed through the amount of calmodulin conjugated Alexa Fluor 488 as fluorescent probes. NCA activity was evaluated by phosphate group released from ATP in enzymatic reaction. The maximum binding capacity (B_{max}) and dissociation constant (K_d) of IP₃R, IP₄R and RyR were determined by radioligand binding of [³H]IP₄, [³H]IP₃ and [³H]-ryanodine to isolated cardiomyocyte nuclei. All results are shown in the table. Our findings suggest that in vivo rat myocardial IRI is characterized by Ca²⁺_n overload, upregulations of nuclear IP₃R and IP₄R, downregulations of NCA activity and RyR, and an increase in permeability of NPC. The upregulation of nuclear IP₃R and IP₄R may be responsible for the nuclear calcium overload.

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ATP-evoked Ca²⁺ waves Stimulate Gene Expression In Human Airway Fibroblasts

Luke Janssen, Yongde Zhang, Tahseen Rahman, Martin Kolb.

McMaster University, Hamilton, ON, Canada.

We sought to investigate the effects of a variety of autacoids on Ca²⁺-handling in human airway fibroblasts. Primary cultured fibroblasts were loaded with the Ca²⁺-indicator dye fluo-4 and studied using confocal fluorimetric microscopy. ATP (10⁻⁵ M) evoked recurring Ca²⁺-waves. This fluorimetric change was

greater and longer lasting within the nucleus of the cell than in the non-nuclear portion of the cytosol, and was only sometimes accompanied by a contraction. These responses were completely occluded by cyclopiazonic acid (10⁻⁵ M; depletes the internal Ca²⁺-store) or the phospholipase C inhibitor U73122 (10⁻⁶ M). Pretreatment of the cells with ryanodine (10⁻⁵ M), on the other hand, had no effect on the ATP-evoked responses. With respect to the receptor through which this response was exerted, we found it to be mimicked by UTP or ADP but not by adenosine or α,β-methylene-ATP, and to be blocked by the purinergic receptor blocker PPADS; interestingly, PPADS itself appeared to sometimes evoke a rise in [Ca²⁺]_i on its own. ATP also evoked a membrane conductance change with characteristics of a non-selective cation current, markedly enhanced synthesis of the cytokine TGFβ and the matrix proteins fibronectin and collagen I; these changes in protein synthesis were blocked by PPADS and were partially reduced by ryanodine. We conclude that, in human pulmonary fibroblasts, ATP acts upon P2Y receptors to liberate internal Ca²⁺ through ryanodine-insensitive channels, leading to a Ca²⁺-wave which courses throughout the cell and triggers protein synthesis.

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Store-operated Ca²⁺ Entry Is Suppressed During Mitosis Due To Phosphorylation Of The Endoplasmic Reticulum Ca²⁺ Sensor Stim1

Jeremy T. Smyth, Rebecca R. Boyles, James W. Putney.

National Institute of Environmental Health Sciences, Research Triangle Park, NC, USA.

When ER Ca²⁺ stores are depleted due to physiological Ca²⁺ release or pharmacological perturbation, Ca²⁺ influx via plasma membrane Ca²⁺ channels is activated by a process known as store-operated Ca²⁺ entry (SOCE). The current associated with SOCE is Ca²⁺ release-activated Ca²⁺ current (I_{crac}). SOCE involves Orai Ca²⁺ influx channels and STIM ER Ca²⁺ sensors. When ER Ca²⁺ stores are full, STIM1 is localized throughout the ER membrane; however, ER Ca²⁺ store depletion induces rearrangement of STIM1 into punctate structures near the plasma membrane where it activates Orai channels. Interestingly, mitosis is the only known physiological situation in which Ca²⁺ store depletion is dissociated from SOCE or I_{crac} activation. Identification of the molecular components of the SOCE signaling pathway has facilitated analysis of the mechanism underlying mitotic SOCE suppression. We found that in mitotic HeLa cells, an enhanced yellow fluorescent protein-tagged STIM1 (eYFP-STIM1) did not rearrange into puncta in response to Ca²⁺ store depletion and accordingly, SOCE was not activated. We hypothesized that mitosis-specific phosphorylation of STIM1 may underlie the block of STIM1 rearrangement and SOCE suppression. To this end, the phospho-specific MPM-2 antibody recognized eYFP-STIM1 immunoprecipitated from mitotic but not interphase cells. MPM-2 recognizes phosphorylated serine or threonine followed by proline, and human STIM1 contains 10 instances of S/T-P, all located in the cytoplasmic, C-terminus. STIM1 truncation mutants indicate that at least 2 sites within the C-terminus account for the mitosis-specific phosphorylation. Individual phosphorylation site mutants are being created to identify specific phosphorylated residues and to determine the functional consequences of phosphorylation during mitosis. Suppression of SOCE during mitosis may be an important signaling event, because mitotic processes such as chromosome separation and cytokinesis are exquisitely sensitive to small changes in cytoplasmic Ca²⁺.

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Heteromeric channel assembly of Orai1 and Orai3 exhibits altered Ca²⁺ selectivity

Rainer Schindl, Irene Frischauf, Judith Bergsmann, Martin Muik,

Isabella Derler, Christoph Romanin.

University of Linz, Linz, Austria.

Coexpression of STIM1, targeted to the endoplasmic reticulum and each of the three Orai (also termed CRACM) channels located in the plasma-membrane leads to store-operated, highly Ca²⁺ selective currents. While Orai1 has been reported to form the native Ca²⁺ release activated Ca²⁺ (CRAC) channels in human T-cells, the molecular architecture of less Ca²⁺ selective store-operated currents remains unknown. Here we show employing confocal fluorescence resonance energy transfer (FRET) that all three Orai proteins are able to form homo- and hetero oligomers. Overexpressed homomeric Orai1 or Orai3 together with STIM1, resulted in store-operated inward rectifying, highly Ca²⁺ selective currents, as resolved by whole-cell patch-clamp recordings. Coexpression of Orai1 together with Orai3 and STIM1 yielded similar store-depletion activated Ca²⁺ currents, yet with a leftward shifted reversal potential, pointing to less selective currents. In line, a tandem construct where Orai1 was linked to Orai3 exhibited a similarly reduced Ca²⁺ selectivity that allowed for robust Cs⁺ permeation. Moreover, Orai3 pore mutants coexpressed with wild-type Orai3 affected Ca²⁺ and Cs⁺ selectivity/permeability. These